

TROPICAL AGRICULTURAL SCIENCE

Journal homepage: http://www.pertanika.upm.edu.my/

Review Article Effects of Spirulina platensis and Chlorella vulgaris on the Immune System and Reproduction of Fish

Wizilla Janti Joshua¹ and Zarirah Zulperi^{1,2*}

¹Aquatic Animal Health and Therapeutics Laboratory, Institute of Bioscience, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia ²Department of Aquaculture, Faculty of Agriculture, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

ABSTRACT

This review briefly highlights previous studies on the effects of *Spirulina platensis* and *Chlorella vulgaris* on the health and reproduction of fish. These microalgae have diverse potentials. This study can be used as a stepping stone in advancing the aquafeed industry by formulating microalgae-based feeds. It can be made to specifically enhanced the health status of fish and its reproductive system through the supplementation and/or replacement of fishmeal or other plant proteins such as soybean meal. Hence, it could be more sustainable than depending on natural fish stocks. The usage of antibiotics and vaccines to solve the issue of disease outbreak in aquaculture, as well as the usage of hormones for the growth and reproduction of fish, can also be replaced by the usage of *S. platensis* and *C. vulgaris*. The inclusion of these microalgae in fish feed has affected hemathological parameters and survival in fish as it boosts the numbers of white and red blood cells and thus affecting the immunity-stimulating capacity in fish. Besides, these microalgae also affect the fecundity and survival of fish eggs production and survival in fish whereas *C. vulgaris* enhances

ARTICLE INFO

Article history: Received: 15 June 2020 Accepted: 10 September 2020 Published: 27 November 2020

DOI: https://doi.org/10.47836/pjtas.43.4.01

E-mail addresses: wizillajoshua@gmail.com (Wizilla Janti Joshua) zarirah@upm.edu.my (Zarirah Zulperi) * Corresponding author oxidative stress that affects the reproduction of White rabbits. This review aimed to deliver the results on the research of *S. platensis* and *C. vulgaris* on the immunity and reproduction of various fish species.

Keywords: *Chlorella vulgaris*, immunity, microalgae, reproduction, *Spirulina platensis*

ISSN: 1511-3701 e-ISSN: 2231-8542

INTRODUCTION

The increasing human population worldwide has pressured the natural stocking of fish. Hence, aquaculture industries are blooming all over the world to support the growing demand for protein sources. This industry also provides food security and directly responsible in developing the livelihood of less-privileged communities. Aquaculture industry is being enforced and promoted as an important tool that drives economic growth in Malaysia and has been enlisted in the National Key Economic Area (NKEA), which highlights the 16 agro-food's entry point projects (EPP) (Yusoff, 2015). Statistics from the Department of Fisheries Malaysia (DOF) (2018) reported that the production of freshwater aquaculture was 102,500 tonnes in 2017 and increased to 105,700 tonnes in 2018, while the production of brackish water aquaculture was 324,300 tonnes in 2017 and 290,900 tonnes in 2018. The rapid growth of the aquaculture industry, particularly fish farming, has raised a number of issues in the health management of fish in terms of fish immunity against disease outbreak and the continuous supply of good quality fish seeds in terms of fish reproduction and developments.

This scenario is often related to the use of chemicals in aquaculture for a successful production (Subasinghe, 2004). However, such application is not widely encouraged as it introduces several risks to the production system, environment, and human health (Melba & Rohana, 2008). Thus, chemotherapeutics has been replaced with other alternative sources that are more acceptable in aquaculture practises, such as adding microalgae in fish feed as supplements. Various forms of algal meal applications have been studied. Generally, microalgae are used as larval feed by some fish farmers, and different levels of inclusion are added in fish feeds to boost their beneficial effects on fish conditions (Brown, 2002). Several microalgae possess high protein, lipid, and carbohydrate contents. Besides, the biomass of microalgae is rich in proteins, and it can strive and compete fairly for its quality and quantity compared with regular food proteins, such as fish, soybeans, and eggs (Ejike et al., 2017). A handful of researches and studies had been carried out in the past to analyse the potential effects of microalgae in fish, and such attempts are still in progress with various developments on specific parameters in fish, such as the immune and health system as well as the reproductive system. In this review, the effects of S. platensis and C. vulgaris on the immunity and reproduction of fish are briefly elaborated.

POTENTIALS OF MICROALGAE IN AQUACULTURE

The growth of the aquaculture industry has gone hand in hand with the growth of the population worldwide to ensure that ample fish supply as a part of protein sources can be fulfilled. The aquaculture industry is expected to solve food security and nutritional well-being, reduce poverty, and develop the economy (Melba & Rohana, 2008). However, the focus on developing

the aquaculture industry to fulfil such expectation has raised several issues in aquaculture. Among the most recognisable problem is the use of chemotherapeutants in aquaculture. Chemotherapeutics has benefits and is a key to a successful aquaculture production in activities, such as pond and tank constructions, feed formulation, growth promotion, health management, and booster of natural production (Melba & Rohana, 2008; Subasinghe, 2004). Nevertheless, the use of chemicals in aquaculture is not widely encouraged. Various researches and tests have been carried out to develop products that are friendlier to replace the use of chemicals in aquaculture. The organism that is known to be a more sustainable choice in replacing the use of chemotherapeutants in aquaculture is microalga. The growing interest in the multipurpose properties of microalgae has developed various microalgae applications in daily life. Microalgae are an alternative to sustainable aquaculture practices by playing roles in wastewater treatment and ingredient replacement in fish feed. The development of fish feed with the inclusion of microalgae is widely studied because of its encouraging effects on farmed fish. Microalgae can be used in aquaculture as feed, growth enhancers, and immunostimulants (Ahmad et al., 2018). However, microalgae must be easily cultured and nontoxic to be used in aquaculture (Spolaore et al., 2006). The criteria that are taken into consideration before incorporating microalgae as feed ingredients for fish are that they need to have the correct size, are easily ingested and digested by fish, and have high nutritional quality profiles (Brown et al., 1999; Renaud et al., 2002). Research on microalgae, such as *S. platensis* and *C. vulgaris*, their effects in enhancing the immunity and reproduction performance of fish and other organisms is continuous and evolving.

GENERAL INTRODUCTION

Spirulina platensis

Spirulina or Arthrospira platensis is a blue-green filamentous alga that inhabits freshwater bodies. Its name is acquired from its cylindrical shape with multicellular trichomes in an open lefthanded helix (Figure 1) (Jung et al., 2019). Morphologically, S. platensis is a helicoid alga with a radial that is distinct from those of other species (Promya et al., 2008). This microalga has a protein content of up to 70% and is rich in vitamins, minerals, and essential fatty acids, such as linolenic and linoleic acids, and palmitic acid (Abdel-Tawwab & Ahmad, 2009). Spirulina has been used as a dietary supplement for a long time by the community that resides near the alkaline lakes where it is habitually found (Jung et al., 2019). Today, Spirulina is produced in numerous countries in Africa and America, such as Benin, Burkina Faso, Chad, Brazil, Chile, and Costa Rica, as well as Asian countries, such as Thailand, India, China, Vietnam, and Taiwan (Habib et al., 2008). However, the production of Spirulina is carried out in control conditions to avoid contaminations from other sources, such as blue-green algae, pesticides or heavy metal; thus, its general composition is affected by

the location and type of production (Table 1) (Jung et al., 2019). The rich contents of this microalgae species made it an interesting ingredient for testing as feed in fish. Various studies have analysed the immunology, disease resistance, oxidative stress, and growth performance of a number of aquatic animals based on *S. platensis*-supplemented feeds (Abdel-Latif & Khalil, 2014; El-Sheekh et al., 2014; Kim et al., 2013; Macias-Sancho et al., 2014; Promya & Chitmanat, 2011; Teimouri et al., 2013; Yeganeh et al., 2015).



Figure 1. The microscopic view of Spirulina Note. Adapted from "Earth food Spirulina (Arthrospira): Production and quality standarts", Retrieved June 01, 2020, from https://www. intechopen.com/books/food-additive/earth-foodspirulina-arthrospira-production-and-qualitystandarts. Copyright 2012 by Koru. Adapted with permission

Table 1

The general composition of Spirulina. Adapted from "Spirulina platensis, a super food?" (Jung et al., 2019)

Components	Percentage (%)
Proteins	55-70
Carbohydrates	15-25
Lipids	6-8
Minerals	7-13
Humidity (dried algae)	3-7
Dietary fibers	8-10

Chlorella vulgaris

Chlorella vulgaris is a freshwater species and unicellular alga that contains a nutrientdense super food, including various vitamins and minerals, 18 amino acids, and 60% protein (Khani et al., 2017). A report by Nick (2003) also stated that Chlorella possessed excess minerals, such as iron, calcium, potassium, magnesium, phosphorous, and 20 vitamins, such as pro-vitamin A, vitamins C, B1, B2, B2, B5, B6, B12, E, K; biotin, inositol, and folic acid. Morphologically, Beijerinck (as cited in Safi et al., 2014, p. 266) discovered C. vulgaris as the first microalga with a well-defined nucleus. Chlorella has a rigid cell wall that varies according to each growth phase and provides protection against invaders and harsh environment (Safi et al., 2014) (Figure 2). In addition,



Figure 2. The growing phases of *Chlorella vulgaris Note*. Adapted from "Morphology, composition, production, processing and applications of *Chlorella vulgaris*: A review" by C. Safi, B. Zebib, O. Merah, P. Y. Pontalier, and C. Vaca-Garcia, 2014, *Renewable and Sustainable Energy Reviews*, *35*, pp. 265–278. Copyright by Elsevier

Pertanika J. Trop. Agric. Sc. 43 (4): 429 - 444 (2020)

this microalga has many fundamental components that are identical to actual plants, including cytoplasm, mitochondrion, and chloroplast. Besides, it is one of the microalgae that contain the highest quantity of chlorophyll compared with other plants (Raji et al., 2018). *Chlorella* has a unique phytonutrient property that is made up of vitamins, nucleic acid-related substances, amino acids, proteins, peptides, and sugars known as the *Chlorella* growth factor (CGF), which is found in abundance in the nuclei of the alga (Nick, 2003).

ENHANCEMENT OF FISH IMMUNE SYSTEM

Spirulina platensis

The properties of S. platensis that are rich in proteins and vitamins have a positive impact on the immunity of the animals that consumed it (Promya & Chitmanat, 2011). However, the effects of S. platensis on the immune system of fish may vary according to the levels of supplementation or inclusion. Promya and Chitmanat (2011) stated that a 5% supplementation of Spirulina in the feed resulted in higher red blood cell (RBC) and white blood cell (WBC) counts in African sharptooth catfish as well as advanced its immunity-stimulating capacity. The result is remarkedly different compared to a 3% supplementation of Spirulina, which gave a lower immunity in African sharptooth catfish. Besides, a feeding experiment on Nile tilapia with five levels of Spirulina inclusion (1.25, 2.5, 5.0, 7.5, and 10.0 g kg⁻¹ diet) added to the basal diet showed that the RBC and WBC counts increased from $1.92\times10^6\,\mu L^{-1}$ to $2.54\times10^6\,\mu L^{-1}$ and 3.21 $\times 10^6 \,\mu\text{L}^{-1}$ to $4.02 \times 10^6 \,\mu\text{L}^{-1}$, respectively, as the Spirulina levels increased, although 5.0 g kg^{-1} Spirulina is the optimum amount that gave the highest specific growth rate, weight gain, and food conversion ratio with no substantial difference in the survival rate from one treatment to another (Abdel-Tawwab & Ahmad, 2009). These results showed that different levels of Spirulina inclusion in fish diet affect blood counts. Blood biochemical indexes can be an indicator that reflects the health status, physiological condition, metabolism, and immunity of fish (Zhou et al., 2001). In addition, alkaline phosphate (ALP) activity in fish is another parameter that is commonly used to study the immunity of fish. ALP is a non-specific phosphate hydrolase that is involved in fish metabolic regulation and plays a vital role in non-specific immune response in organisms (Huang et al., 2005). Lin et al. (2016) showed an increasing trend in the ALP activity of golden pomfret (Trachinotus ovatus) based on the 0%-6% supplementation of S. platensis in the fish diet and suggested that further experiment should be carried out for the ALP activity in fish. C-phycocyanin, a property of Spirulina, helps build immune capacity (Vonshak, 1997) and could be the reason for the enhancement of the immunity of fish based on the feeding trials with the inclusion or supplementation of Spirulina. Furthermore, a different approach of administering Spirulina through direct or oral method was

also investigated. A test on carp (Cyprinus caprio) intubated with 0 (control), 1, 10, and 25 mg doses of Spirulina suspended in sterilised physiological saline (0.85% NaCl) and subjected to Aeromonas hydrophilia (strain MU9901) infection showed that the number of bacterial cells in the liver and kidney of carp treated with Spirulina were lower compared with those of the control groups at the treatment intervals of 4, 8, and 12 hours and at 1 and 4 hours post-bacterial challenge (Watanuki et al., 2006). Another challenge test using sturgeon (Huso huso) that was intraperitoneally injected with Streptococcus iniae (strain ATCC29178) after 8 weeks of feeding period with 0%, 2.5%, 5%, and 10% of Spirulina added to the basal diet showed that the cumulative mortality of sturgeon decreased with the increasing supplementation of Spirulina (Adel et al., 2016). These challenge tests suggested that Spirulina could activate leucocyte activities in fish and thus increased the resistance against bacterial infections. Leucocytes are involved in superoxide production, cytokine release, and phagocytosis (Watanuki et al., 2006). The effects of Spirulina in enhancing the immunity of fish from disease infections and increasing the blood biochemical indexes in fish could create a specific market for Spirulina-based feed or supplements for aquaculture use. Spirulina could also be an alternative to replace the usage of antibiotics and vaccines, which are less environmentally friendly and have higher cost.

Chlorella vulgaris

Chlorella vulgaris is a green microalgae species with an engaging immunostimulant property that enhances the health and increases the life expectancy of fish (Gouveia et al., 2002). Formulated diet supplemented with 2%, 5%, 7%, and 10% of C. vulgaris dry powder gave higher values of C4, total immunoglobulin and lysozyme in koi carp than the control group (0% Chlorella vulgaris inclusion); thus, C. vulgaris could be involved in the modulation of the innate immunity of fish during the experimental period of 8 weeks (Khani et al., 2017). Khani et al. (2017) also found that the koi group fed with C. vulgaris-supplemented diets also had higher haemoglobin and haematocrit levels, and the highest values were obtained by 5% Chlorella vulgaris supplementation. These components are important for the survival of fish and linked to the oxygenbinding capacity of blood (Bielek & Strauss, 1993). The addition of C. vulgaris powder supplement to the formulated diet increased the levels of IgM, IgD, interleukin-22 (IL)-22, and chemokine (C-C motif) ligand 5 in Gibel carp (Carassius auratus gibelio) (Zhang et al., 2014). The expression of various cytokines, such as IL-8 and IL-1, in fish fed with microalga-supplemented diet is greatly affected (Díaz-Rosales et al., 2008). This report supports the immunostimulant potential of C. vulgaris in boosting the immunity of fish. Besides, C. vulgaris contains abundant carotenoid and thus subjected to various studies in yielding different types of carotenoids (Markou & Nerantzis, 2013). Fish with

high carotenoid content are more immune to fungal and bacterial infections (Gupta et al., 2007). Besides, carotenoids are soluble lipid pigments that play a role in the formation of skin colouring in ornamental fish [Kestemont et al. (as cited in Liang et al., 2012, p. 2); Paripatananont et al., 1999]. The biomass of C. vulgaris is more efficient in enhancing skin colouring and produces the highest deposition of carotenoid and red hue in three varieties of chromatic koi carps, namely, Kawari (red), Showa (black and red), and Bekko (black and white) (Gouveia et al., 2003). Colouring and pattern formation are important in ornamental fish as those traits will determine their quality (Li et al., 2008). The high bioavailability of carotenoids in C. vulgaris and the thin cellular membrane of this microalga are the possible characteristics that gave better efficiency for the skin colouring of fish, even for species that have relatively short digestive tracts (Gouveia et al., 1998). Chlorella vulgaris also remarkably enhances the growth performance and feed intake of koi carp at 5% inclusion in feed and other parameters, such as specific growth rate and protein and lipid efficiency rate, in contrast to the group without any C. vulgaris inclusion; thus, the growth enhancement is possibly caused by the high digestibility of this microalga (Khani et al., 2017). This enhancement may also be due to growth promoter properties, such as the adequate amounts of macronutrient and CGF in C. vulgaris (Badwy et al., 2008; Yamaguchi, 1996) and its highquality protein content (Kang et al., 2013).

However, further research on the growth effects of *Chlorella* should be carried out in various fish species and might not give the same positive effects. Its effect can be affected by the levels of supplementation, species-specific reactions to *Chlorella*, and the experimental period (Rahimnejad et al., 2016). Hence, the addition of *C. vulgaris* in feed for its carotenoid contents and property in stimulating the growth and immunity of fish is a promising idea as *C. vulgaris* is also known as a fast-growing microalga and can be a sustainable aquafeed ingredient in the future.

The addition or inclusion of *S. platensis* and *C. vulgaris* in fish diet and their effects on fish immune system are summarised in Table 2. Further trials on these microalga species on different fish stages, sizes, age, and environment should be investigated to improve the knowledge of suitable fish diet formulation with the addition of microalgae as natural supplements that could boost the immune system of fish. However, such approach requires a broad understanding on the abilities of different fish species to digest or utilise the target properties in microalgae, particularly the immunostimulant properties.

IMPROVEMENT OF FISH REPRODUCTION SYSTEM

Spirulina platensis

The effects of *S. platensis* on the reproduction and survival rates of fish were studied by several researchers throughout the years. The quality of fish brooders is the important key to a successful fish propagation. The features that define the reproductive capability and

Wizilla Janti Joshua and Zarirah Zulperi

Table 2

Summary of addition or inclusion of Spirulina platensis and Chlorella vulgaris in fish diet and their effects on fish immunity

Microalgae	Fish species	Diet / Test type(s)	Effects on immunity	Reference(s)
	<i>Clarias</i> <i>gariepinus</i> (African sharptooth catfish)	D1 (Control): Instant fish feed (obtained from animal feed supply) D2: Instant fish feed + 3% <i>Spirulina</i> algae D3: Instant fish feed + 5% <i>Spirulina</i> algae	Higher counts of WBC, RBC, and lysozyme activity assay in 3% and 5% <i>Spirulina</i> supplementation diets as compared to control	Promya and Chitmanat (2011)
	Oreochromis niloticus, L. (Nile tilapia)	D1 (Control): Basal diet (30.6% crude protein, 9.1% lipids, 4.72 kcal GE g ⁻¹) D2: Basal diet + 1.25 g <i>Spirulina/</i> kg diet D3: Basal diet + 2.5 g <i>Spirulina/</i> kg diet D4: Basal diet + 5.0 g <i>Spirulina/</i> kg diet D5: Basal diet + 7.5 g <i>Spirulina/</i> kg diet D6: Basal diet + 10.0 g <i>Spirulina/</i> kg diet	Increasing number of WBC, RBC, and lymphocytes as the amount of <i>Spirulina</i> added in diets increased	Abdel- Tawwab and Ahmad (2019)
*	<i>Trachinotus</i> <i>ovatus</i> (Golden pomfret)	D1 (Control): Basal diet (50.27% crude protein, 8.35 crude lipid) D2: Basal diet + 1.0% <i>Spirulina</i> D3: Basal diet + 2.0% <i>Spirulina</i> D4: Basal diet + 3.0% <i>Spirulina</i> D5: Basal diet + 4.0% <i>Spirulina</i> D6: Basal diet + 5.0% <i>Spirulina</i>	Increasing trend in the alkaline phosphate (ALP) activity as the amount of <i>Spirulina</i> in the diet increased	Lin et al. (2016)
	<i>Cyprinus</i> <i>caprio</i> (Carp)	Direct intubation (0.1 ml suspension) Dose 1: 0 dose (0.85% NaCl) Dose 2: 0.85% NaCl + 1 mg <i>Spirulina</i> Dose 3: 0.85% NaCl + 10 mg <i>Spirulina</i> Dose 4: 0.85% NaCl + 25 mg <i>Spirulina</i>	Lower number of bacterial cell (challenged with <i>Aeromonas</i> <i>hydrophilia</i> - strain MU9901 infection) in liver and kidney as compared to control	Watanuki et al. (2006)
	Huso huso (Sturgeon)	D1: Basal diet (Control) D2: Basal diet + 2.5% pure dried <i>Spirulina</i> D3: Basal diet + 5.0% pure dried <i>Spirulina</i> D4: Basal diet + 10.0% pure dried <i>Spirulina</i>	Cumulative mortality of the sturgeon decreased with the increasing supplementation of the <i>Spirulina</i> in basal diet (challenged with <i>Streptococcus iniae</i> - strain ATCC29178) through intraperitoneal injection	Adel et al. (2016)

Spirulina platensis and Chlorella vulgaris as Feed Supplements

Table 2 ((continue)	
14010 2 (continue	

Microalgae	Fish species	Diet / Test type(s)	Effects on immunity	Reference(s)
	<i>Cyprinus</i> <i>caprio</i> (Carp)	D1: Formulated feed (Control) D2: Formulated feed + 2% dry powder <i>C. vulgaris</i> D3: Formulated feed + 5% dry powder <i>C. vulgaris</i> D4: Formulated feed + 7% dry	Higher values of immune parameters: C4, total immunoglobulin, and lysozyme than the control	Khani et al. (2017)
Chlorella vulgaris		powder <i>C. vulgaris</i> D5: Formulated feed + 10% dry powder <i>C. vulgaris</i>	Higher values of haemoglobin (Hb) and haematocrit (Ht) than control – 5% of <i>Chlorella vulgaris</i> supplementation gave highest values	
2	Carassius auratus gibelio (Gibel carp)	D1: Formulated diet (Control) D2: Formulated diet + 0.4% <i>Chlorella</i> powder D3: Formulated diet + 0.8% <i>Chlorella</i> powder D4: Formulated diet + 1.2% <i>Chlorella</i> powder D5: Formulated diet + 1.6% <i>Chlorella</i> powder D6: Formulated diet + 2.0% <i>Chlorella</i> powder	Increasing immunoglobulin (Ig) M and D, interleukin-22 (IL)-22, and chemokine (C-C motif) ligand 5 (CCL-5) as the <i>Chlorella</i> supplementation in diet increased	Zhang et al. (2014)

performance of brooders rely on the eggs' fecundity, diameter, and hatchability rates (Chong et al., 2004; Kumaraguruvasagam et al., 2007; Izquierdo et al., 2001). Spirulina platensis provides a remarkably higher egg survival rate of 73%, shorter hatching and faster larval development compared with the commercial flake diet-fed group when given as sole food for zebrafish broodstock with the hypothesis that commercial flake does not contain as much Omega-6 fatty acid as S. platensis (Geffroy & Simon, 2013). In addition, S. platensis contains omega-6 fatty acid (which made up 41.2% of fatty acids), specifically gamma linolenic acid and linoleic acid (Qiang et al., 1997). These properties are the precursors of arachidonic acid, which is a remarkable constituent in the formation of prostaglandin that interferes in oocyte maturation, ovulation, and steroidogenesis (Pati & Habibi, 2002; Patiño & Sullivan, 2002). Hence, Spirulina is a promising diet for adult fish. Spirulina platensis also considerably increases the hatching percentage and total egg reproduced by yellow tail cichlid (Pseudotropheus acei) during a 12-week observation period (Güroy et al., 2012). This microalga species could possibly replace the administration of artificial hormones in enhancing reproduction performance by incorporating it in the diet of brooders. Besides, raw Spirulina was also tested on Nile tilapia as the primary feed and has increased

the egg production, hatching percentage, and survival rates of fish compared with conventional fish feed (Promya & Chitmanat, 2011). Spirulina as a replacement fish meal for the feeding of three-spot gourami (Trichopodus trichopterus) provides greater gonadosomatic indices (19.4%-21.85%) and affects the absolute fecundity of fish between 7,300 and 12,700 eggs per female at 2.5%-10% Spirulina platensis replacement levels than the group fed with fish meal only (Khanzadeh et al., 2016). The addition of this microalga can result in the enhancement of gonad maturation. Apart from that, S. platensis possesses fat soluble pigments (carotenoids), such as xanthophylls, B-carotene, echinenone, cryptoxanthin, and zeaxanthin (Nakagawa & Montgomery, 2007). Reproductive performance can be enhanced by dietary carotenoids (Watanabe & Vassalo-Agius, 2003). Increasing dietary carotenoid supplementation enhances the reproduction capability of different fish species, such as rainbow trout (Oncorhynchus mykiss) (Dabrowski et al., 1987), gilthead seabream (Sparus aurata) (Scabini et al., 2010), and yellow tail (Seriola quinqueradiata) (Vassallo-Agius et al., 2001, 2002). These discoveries show that the carotenoids in S. platensis play multiple roles, that is, it affects the colouration of fish and improves their reproductive performance. Thus, S. platensis can be one of the primary sources to be developed in ornamental fish industry by focusing on its capability to enhance the colouration and reproductive performance of the fish.

Chlorella vulgaris

Based on the knowledge obtained in this review, very scarce information is available on the effects of C. vulgaris on the reproduction of fish. Some of the studies found were only focused on mammals. For example, Sikiru et al. (2019) reported that C. vulgaris enhances oxidative stress, which was an exclusive biochemical complication that affected the reproduction in New Zealand White rabbits. Besides, the extract of C. vulgaris improves the histological adjustment of monosodium glutamate in ovarian tissue, the level of sex hormones, and increases the level of ovarian enzymatic antioxidants in adult female albino mice (Abdel-Aziem et al., 2018). Hence, its effects on fish should be further explored for potential use in aquaculture.

FUTURE PROSPECT AND CONCLUSION

The inclusion of Spirulina platensis and Chlorella vulgaris as feed supplement has substantial impacts on fish health and immunity. Planning better tests on a bigger scale of fish culture system is important to see the pattern of immunity and hematological parameters in natural culture conditions than in controlled laboratory environments. Besides, S. platensis has a potential in affecting the performance of fish brooder, which directly influences the quality and survival of eggs. Well-modified feed products based on S. platensis that focus more on the reproductive effects in fish could have a great future in aquaculture. Apart from that, more research should be

done to analyse the effects of C. vulgaris on fish to highlight the capability of carotenoids in evaluating the growth performance and immunity of fish. In conclusion, this review briefly presented the effects of S. platensis and C. vulgaris on the immunity and reproduction of fish. It is recommended that more studies should be conducted on the possible contributions of these microalgae in the aquafeed industry to create more environmentally effective antibiotics and vaccines to combat fish diseases in a culture system and enhance the reproduction of fish. Using renewable natural products, such as microalgae, will also benefit the aquaculture industry by having better practices that conserve the environment.

ACKNOWLEDGEMENT

This study was funded by the Ministry of Higher Education Malaysia through the SATREPS-COSMOS project (JPMJSA 1509).

REFERENCES

- Abdel-Aziem, S., Abd El-Kader, H., Ibrahim, F., Sharaf, H., & El Makawy, A. (2018). Evaluation of the alleviative role of *Chlorella vulgaris* and *Spirulina platensis* extract against ovarian dysfunctions induced by monosodium glutamate in mice. *Journal of Genetic Engineering and Biotechnology*, *16*(2), 653-660. doi: 10.1016/j. jgeb.2018.05.001
- Abdel-Latif, H. M. R., & Khalil, R. H. (2014). Evaluation of two phytobiotics, Spirulina platensis and Origanum vulgare extract on growth, serum antioxidant activities and resistance of Nile tilapia (Oreochromis niloticus) to pathogenic Vibrio alginolyticus. International

Journal of Fisheries and Aquatic Studies, *1*(5), 250–255.

- Abdel-Tawwab, M., & Ahmad, M. H. (2009). Live Spirulina (Arthrospira platensis) as a growth and immunity promoter for Nile tilapia, Oreochromis niloticus (L.), challenged with pathogenic Aeromonas hydrophila. Aquaculture Research, 40(9), 1037-1046. doi: 10.1111/j.1365-2109.2009.02195.x
- Adel, M., Yeganeh, S., Dadar, M., Sakai, M., & Dawood, M. A. O. (2016). Effects of dietary *Spirulina platensis* on growth performance, humoral and mucosal immune responses and disease resistance in juvenile great sturgeon (*Huso huso* Linnaeus, 1754). *Fish and Shellfish Immunology*, 56, 436-444. doi: 10.1016/j. fsi.2016.08.003
- Ahmad, M. T., Shariff, M., Md. Yusoff, F., Goh, Y. M., & Banerjee, S. (2020). Applications of microalga *Chlorella vulgaris* in aquaculture. *Reviews in Aquaculture*, 12(1), 328-346. doi: 10.1111/raq.12320
- Badwy, E. M., Ibrahim, M. M., & Zeinhom, M. M. (2008). Partial replacement of fishmeal with dried microalga (*Chlorella* spp. and *Scenedesmus* spp.) in Nile tilapia (*Oreochromis niloticus*) diets. In 8th International Symposium on Tilapia in Aquaculture (pp. 801–811). Cairo, Egypt: Ministry of Agriculture.
- Bielek, E., & Strauss, B. (1993). Ultrastructure of the granulocytes of the South American lungfish, *Lepidosiren paradoxa*: Morphogenesis and comparison to other leucocystes. *Journal of Morphology*, 281(1), 29–41. doi: 10.1002/ jmor.1052180103
- Brown, M. R. (2002). *Nutritional value and use* of microalgae in aquaculture. Retrieved June 02, 2020, from https://www.uanl.mx/utilerias/ nutricion acuicola/VI/archivos/A19.pdf
- Brown, M. R., Mular, M., Miller, I., Farmer, C., & Trenerry, C. (1999). The vitamin content

of microalgae used in aquaculture. *Journal* of Applied Phycology, 11(3), 247–255. doi: 10.1023/A:1008075903578

- Chong, A. S. C., Ishak, S. D., Osman, Z., & Hashim, R. (2004). Effect of dietary protein level on the reproductive performance of female swordtails, *Xiphophorus helleri* (Poeciliidae). *Aquaculture*, 234(1–4), 381–392. doi: 10.1016/j. aquaculture.2003.12.003
- Dabrowski, K., Luczynski, M., Czeczuga, B., & Falkowski, S. (1987). Relationships among coregond fish reproductive effort, carotenoid content in eggs and survival of embryos. Archiv für Hydrobiologie. Supplementband. Monographische Beiträge, 79(1), 29-48.
- Department of Fisheries Malaysia. (2018). Annual fisheries statistics. Retrieved June 02, 2020, from https://dosm.gov.my/v1/index.php?r=column/ pdfPrev&id=UjYxeDNkZ0xOUjhFeHpna20w UUJOUT09
- Díaz-Rosales, P., Chabrillón, M., Abdala, R. T., Figueroa, F. L., Balebona, M. C., & Moriñigo, M. A. (2008). Effect of dietary administration of *Porphyridium cruentum* on the respiratory burst activity of sole, *Solea senegalensis* (Kaup), phagocytes. *Journal of Fish Diseases*, *31*(7), 489– 495. doi: 10.1111/j.1365-2761.2008.00923.x
- Ejike, C., Collins, S. A., Balasuriya, N., Andrew, K. S., Mason, B., & Chibuike, C. U. (2017). Prospects of microalgae proteins in producing peptide-based functional foods for promoting cardiovascular health. *Trends in Food Science and Technology*, 59, 30–36. doi: 10.1016/j. tifs.2016.10.026
- El-Sheekh, M., El-Shourbagy, I., Shalaby, I., & Hosny, S. (2014). Effect of feeding *Arthrospira platensis* (*Spirulina*) on growth and carcass composition of hybrid red tilapia (*Oreochromis niloticus* x *Oreochromis mossambicus*). *Turkish Journal of Fisheries and Aquatic Sciences*, 14, 471–478. doi: 10.4194/1303-2712-v14_2_18

- Geffroy, B., & Simon, O. (2013). Effects of a Spirulina platensis-based diet on zebrafish female reproductive performance and larval survival rate. Cybium, 37(1–2), 31–38. doi: 10.26028/cybium/2013-371-004
- Gouveia, L., Choubert, G., Gomes, E., Pereira, N., Santinha, J., & Empis, J. (2002). Colouringation of gilthead seabream, *Sparus aurata* (Lin 1875), using *Chlorella vulgaris* microalga. *Aquaculture Research*, 33(12), 987–993. doi: 10.1046/j.1365-2109.2002.00751.x
- Gouveia, L., Choubert, G., Gomes, E., Rema, P., & Empis, J. (1998). Use of *Chlorella vulgaris* as a carotenoid source for salmonids: Effect of dietary lipid content on colouringation, digestibility and muscular retention. *Aquaculture International*, *6*, 269–279.
- Gouveia, L., Rema, P., Pereira, O., & Empis, J. (2003).
 Colouring ornamental fish (*Cyprinus carpio* and *Carassius auratus*) with microalgal biomass. *Aquaculture Nutrition*, 9(2), 123–129. doi: 10.1046/j.1365-2095.2003.00233.x
- Gupta, S. K., Jha, A. K., Pal, A. K., & Venkateshwarlu, G. (2007). Use of natural carotenoids for pigmentation in fishes. *Natural Products and Radiance*, 6(1), 46–49.
- Güroy, B., Sahin, I., Mantoglu, S., & Kayali, S. (2012). Spirulina as a natural carotenoid source on growth, pigmentation and reproductive performance of yellow tail cichlid Pseudotropheus acei. Aquaculture International, 20(5), 869–878. doi: 10.1007/s10499-012-9512-x
- Habib, M. A. B., Parvin, M., Huntington, T. C. H., & Hasan, M. R. (2008). A review on culture, production and use of Spirulina as food for humans and feeds for domestic animals and fish. Rome, Italy: Food and Agriculture Organization of The United Nations (FAO).
- Huang, Z. Y., Chen, Y. X., Zhao, Y., Zuo, Z. H., Chen, M., & Wang, C. G. (2005). Effects of tributyltin

on alkaline phosphatase, aaciphosphatase andadenosine triphosphatase gill activity of digestive gland in clam *Meretrix meretrix*. *Marine Environmental Science*, 24(3), 56–59.

- Izquierdo, M., Fernandez-Palacios, H., & Tacon, A. G. J. (2001). Effect of broodstock nutrition on reproductive performance of fish. *Aquaculture*, 197(1–4), 25–42. doi: 10.1016/s0044-8486(01)00581-6
- Jung, F., Krüger-Genge, A., Waldeck, P., & Küpper, J. H. (2019). Spirulina platensis, a super food?. Journal of Cellular Biotechnology, 5(1), 43-54. doi: 10.3233/jcb-189012
- Kang, H. K., Salim, H. M., Akter, N., Kim, D. W., Bang, T. H., Kim, M. J., ... Suh, O. S. (2013). Effect of various forms of dietary *Chlorella* supplementation on growth performance, immune characteristics, and intestinal microflora population of broiler chickens. *Journal of Applied Poultry Research*, 22(1), 100–108. doi: 10.3382/japr.2012-00622
- Khani, M., Soltani, M., Shamsaie, M., Foroudi, F., & Ghaeni, M. (2017). The effect of *Chlorella vulgaris* (Chlorophyta, Volvocales) microalga on some hematological and immune system parameters of Koi carp (*Cyprinus carpio*). *Iranian Journal of Ichthyology*, 4(1), 62–68.
- Khanzadeh, M., Fereidouni, A. E., & Berenjestanaki, S. S. (2016). Effects of partial replacement of fish meal with *Spirulina platensis* meal in practical diets on growth, survival, body composition and reproductive performance of three-spot gourami (*Trichopodus trichopterus*) (Pallas, 1770). *Aquaculture International*, 24(1), 69–84. doi: 10.1007/s10499-015-9909-4
- Kim, S. S., Rahimnejad, S., Kim, K. W., & Lee, K. J. (2013). Partial replacement of fish meal with Spirulina pacifica in diets for parrot fish (Oplegnathus fasciatus). Turkish Journal of Fisheries and Aquatic Sciences, 13, 197–204. doi: 10.4194/1303-2712-v13 2 01

- Koru, E. (2012). Earth food Spirulina (Arthrospira): Production and quality standarts. Retrieved June 01, 2020, from https://www.intechopen. com/books/food-additive/earth-food-spirulinaarthrospira-production-and-quality-standarts
- Kumaraguruvasagam, K. P., Shanmugam, A., & Rajagopalan, S. (2007). Dietary effect on fry production and growth performance of sail fin molly, *Poecilia latipinna*, in saltwater. *Acta Ichthyologica Et Piscatoria*, 37(1), 29–35. doi: 10.3750/aip2007.37.1.05
- Li, X. H., Wang, X. J., & Mu, X. D. (2008). The research development of ornamental fish pigmentation. *Hebei Fisheries*, *12*, 6–14.
- Liang, Y. J., Bai, D. Q., Yang, G., Wei, D., Guo, M., Yan, S. S., ... Ning, B. (2012). Effect of astacin on growth and color formation of juvenile redwhite ornamental carp (Cyprinus carpio var. Koi L.). Retrieved June 01, 2020, from https://evols. library.manoa.hawaii.edu/handle/10524/23596
- Lin, H., Chen, X., Yang, Y., Wang, J., Huang, X., Huang, Z., ... Qi, C. (2016). Effect of different levels of *Spirulina platensis* dietary supplementation on the growth, body color, digestion, and immunity of *Trachinotus ovatus*. *Israeli Journal of Aquaculture - Bamidgeh*, 68, 1-9.
- Macias-Sancho, J., Poersch, L. H., Bauer, W., Romano, L. A., Wasielesky, W., & Tesser, M. B. (2014).
 Fishmeal substitution with *Arthrospira (Spirulina platensis)* in a practical diet for *Litopenaeus vannamei*: Effects on growth and immunological parameters. *Aquaculture*, 426, 120–125. doi: 10.1016/j.aquaculture.2014.01.028
- Markou, G., & Nerantzis, E. (2013). Microalgae for high-value compounds and biofuels production: A review with focus on cultivation under stress conditions. *Biotechnology Advances*, 31(8), 1532–1542. doi: 10.1016/j. biotechadv.2013.07.011

- Melba G. B., & Rohana P. S. (2008). Meeting the future demand for aquatic food through aquaculture: The role of aquatic animal health. Rome, Italy: Food and Agriculture Organization of the United Nations (FAO).
- Nakagawa, H., & Montgomery, W. L. (2007). Algae. In H. Nakagawa, M. Sato, & D. M. Gatlin III (Eds.), *Dietary supplements for the health and quality of cultured fish* (pp. 133–167). Wallingford, United Kingdom: Centre for Agriculture and Bioscience International (CABI).
- Nick, G. L. (2003). Addressing human exposure to environmental toxins with Chlorella pyrenoidosamedicinal properties in whole foods. Retrieved June 01, 2020, from https://go.gale.com/ps/ anonymous?id=GALE%7CA99164820&sid= google Scholar&v=2.1&it=r&linkaccess=fulltext &issn=15254283&p=AONE&sw=w
- Paripatananont, T., Tangtrongpairoj, J., Sailasuta, A., & Chansue, N. (1999). Effect of astaxanthin on the pigmentation of goldfish *Carassius auratus*. *Journal of the World Aquaculture Society*, 30(4), 454-460. doi: 10.1111/j.1749-7345.1999. tb00993.x
- Pati, D., & Habibi, H. R. (2002). Involvement of protein kinase C and arachidonic acid pathways in the gonadotropin-releasing hormone regulation of oocyte meiosis and follicular steroidogenesis in the goldfish ovary. *Biology* of Reproduction, 66(3), 813–822. doi: 10.1095/ biolreprod66.3.813
- Patiño, R., & Sullivan, C. V. (2002). Ovarian follicle growth, maturation, and ovulation in teleost fish. *Fish Physiology and Biochemistry*, 26, 57–70. doi: 10.1023/A:1023311613987
- Promya, J., & Chitmanat, C. (2011). The effects of Spirulina platensis and Cladophora algae on the growth performance, meat quality and immunity stimulating capacity of the African sharptooth catfish (Clarias gariepinus). International Journal of Agriculture Biology, 13(1), 77–82.

- Promya, J., Traichaiyaporn, S., & Deming, R. L. (2008). The optimum N: P ratio of kitchen wastewater and oil-extracted fermented soybean water for cultivation of *Spirulina platensis*: Pigment content and biomass production. *International Journal of Agriculture Biology*, 10(4), 437–441.
- Qiang, H., Zhengyu, H., Cohen, Z., & Richmond, A. (1997). Enhancement of eicosapentaenoic acid (EPA) and gammalinolenic acid (GLA) production by manipulating algal density of outdoor cultures of *Monodus subterraneus* (Eustigmatophyta) and *Spirulina platensis* (Cyanobacteria). *European Journal of Phycology*, 32(1), 81–86. doi: 10.1080/09541449710001719395
- Rahimnejad, S., Lee, S. M., Park, H. G., & Choi, J. (2016). Effects of dietary inclusion of *Chlorella vulgaris* on growth, blood biochemical parameters and antioxidant enzyme activity in olive flounder, *Paralichthys olivaceus. Journal of the World Aquaculture Society*, 48(1), 103-112. doi: 10.1111/jwas.12320
- Raji, A. A., Alaba, P. A., Yusuf, I., Bakar, N. H. A., Taufek, N. H. M., Muin, H., ... Razak, S. A. (2018). Fishmeal replacement with *Spirulina platensis* and *Chlorella vulgaris* in African catfish (*Clarias gariepinus*) diet: Effect on antioxidant enzyme activities and haematological parameters. *Research in Veterinary Science*, 119, 67–75. doi: 10.1016/j.rvsc.2018.05.013
- Renaud, S. M., Thinh, L.-V., Lambrinidis, G., & Parry, D. L. (2002). Effect of temperature on growth, chemical composition and fatty acid composition of tropical Australian microalgae grown in batch cultures. *Aquaculture*, 211(1-4), 195–214. doi: 10.1016/s0044-8486(01)00875-4
- Safi, C., Zebib, B., Merah, O., Pontalier, P. Y., & Vaca-Garcia, C. (2014). Morphology, composition, production, processing and applications of *Chlorella vulgaris*: A review. *Renewable and Sustainable Energy Reviews*, 35, 265–278. doi: 10.1016/j.rser.2014.04.007

- Scabini, V., Fernandez-Palacios, H., Robaina, L., Kalinowski, T., & Izquierdo, M. S. (2010). Reproductive performance of gilthead seabream (*Sparus aurata* L., 1758) fed two combined levels of carotenoids from paprika oleoresin and essential fatty acids. *Aquaculture Nutrition*, 17(3), 304–312. doi: 10.1111/j.1365-2095.2010.00766.x
- Sikiru, A. B., Arangasamy, A., Alemede, I. C., Guvvala, P. R., Egena, S. S. A., Ippala, J. R., & Bhatta, R. (2019). *Chlorella vulgaris* supplementation effects on performances, oxidative stress and antioxidant genes expression in liver and ovaries of New Zealand White rabbits. *Heliyon*, 5(9), e02470. doi: 10.1016/j. heliyon.2019.e02470
- Spolaore, P, Joannis-Cassan, C., Duran, E., & Isambert, A. (2006). Commercial applications of microalgae. *Journal of Bioscience and Bioengineering*, 101(2), 87–96. doi: 10.1263/ jbb.101.87
- Subasinghe, R. P. (2004). Risks of chemical usage in aquaculture. In J. R. Arthur & M. G. Bondad-Reantaso (Eds.), Capacity and Awareness Building on Import Risk Analysis for Aquatic Animals: Proceedings of the Workshops in Bangkok, Thailand (2000) and Mazatlan, Mexico (2002) (pp. 33-36). Bangkok, Thailand: Network of Aquaculture Centres in Asia-Pacific (NACA).
- Teimouri, M., Amirkolaie, A. K., & Yeganeh, S. (2013). The effects of *Spirulina platensis* meal as a feed supplement on growth performance and pigmentation of rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*, 396, 14–19. doi: 10.1016/j. aquaculture.2013.02.009
- Vassallo-Agius, R., Watanabe, T., Imaizumi, H., & Yamazaki, T. (2002). Spawning performance of yellowtail *Seriola quinqueradiata* fed dry pellets containing paprika and squid meal. *Fisheries Science*, 68(1), 230–232. doi: 10.1046/j.1444-2906.2002.00414.x

- Vassallo-Agius, R., Watanabe, T., Satoh, S., Kiron, V., Imaizumi, H., Yamazaki, T., & Kawano, K. (2001). Supplementation of paprika as a carotenoid source in soft-dry pellets for broodstock yellowtail *Seriola quinqueradiata* (Temminck & Schlegel). *Aquaculture Research*, 32(S1), 263–272. doi: 10.1046/j.1355-557x.2001.00023.x
- Vonshak, A. (Ed.) (1997). Spirulina platensis (Arthospira): Physiology, cell biology and biotechnology. London, England: Taylor and Francis Ltd.
- Watanabe, T., & Vassallo-Agius, R. (2003). Broodstock nutrition research on marine finfish in Japan. Aquaculture, 227(1–4), 35–61. doi: 10.1016/s0044-8486(03)00494-0
- Watanuki, H., Ota, K., Tassakka, A., Kato, T., & Sakai, M. (2006). Immunostimulant effects of dietary *Spirulina platensis* on carp, *Cyprinus carpio*. *Aquaculture*, 258(1–4), 157–163. doi: 10.1016/j. aquaculture.2006.05.003
- Yamaguchi, K. (1996). Recent advances in microalgal bioscience in Japan, with special reference to utilization of biomass and metabolites: A review. *Journal of Applied Phycology*, 8(6), 487–502. doi: 10.1007/bf02186327
- Yeganeh, S., Teimouri, M., & Amirkolaie, A. K. (2015). Dietary effects of *Spirulina platensis* on hematological and serum biochemical parameters of rainbow trout (*Oncorhynchus mykiss*). *Research in Veterinary Science*, 101, 84–88. doi: 10.1016/j.rvsc.2015.06.002
- Yusoff, A. (2015). Status of resource management and aquaculture in Malaysia. In M. R. R. Romana-Eguia, F. D. Parado-Estepa, N. D. Salayo, & M. J. H. Lebata-Ramos (Eds.), Resource Enhancement and Sustainable Aquaculture Practices in Southeast Asia: Challenges in Responsible Production of Aquatic Species: Proceedings of the International Workshop on Resource Enhancement and Sustainable

Aquaculture Practices in Southeast Asia 2014 (RESA) (pp. 53-65). Tigbauan, Philippines: Aquaculture Department, Southeast Asian Fisheries Development Center.

Zhang, Q., Qiu, M., Xu, W., Gao, Z., Shao, R., & Qi,Z. (2014). Effects of dietary administration of *Chlorella* on the immune status of Gibel carp,

Carassius auratus gibelio. Italian Journal of Animal Science, *13*(3), 3168. doi: 10.4081/ ijas.2014.3168

Zhou, Y., Guo, W. C., Yang, Z. G., & Zhang, K. (2001). Advances in the study of haemotological indices of fish. *Journal of Shanghai Fisheries University*, 10(2), 163-165.